

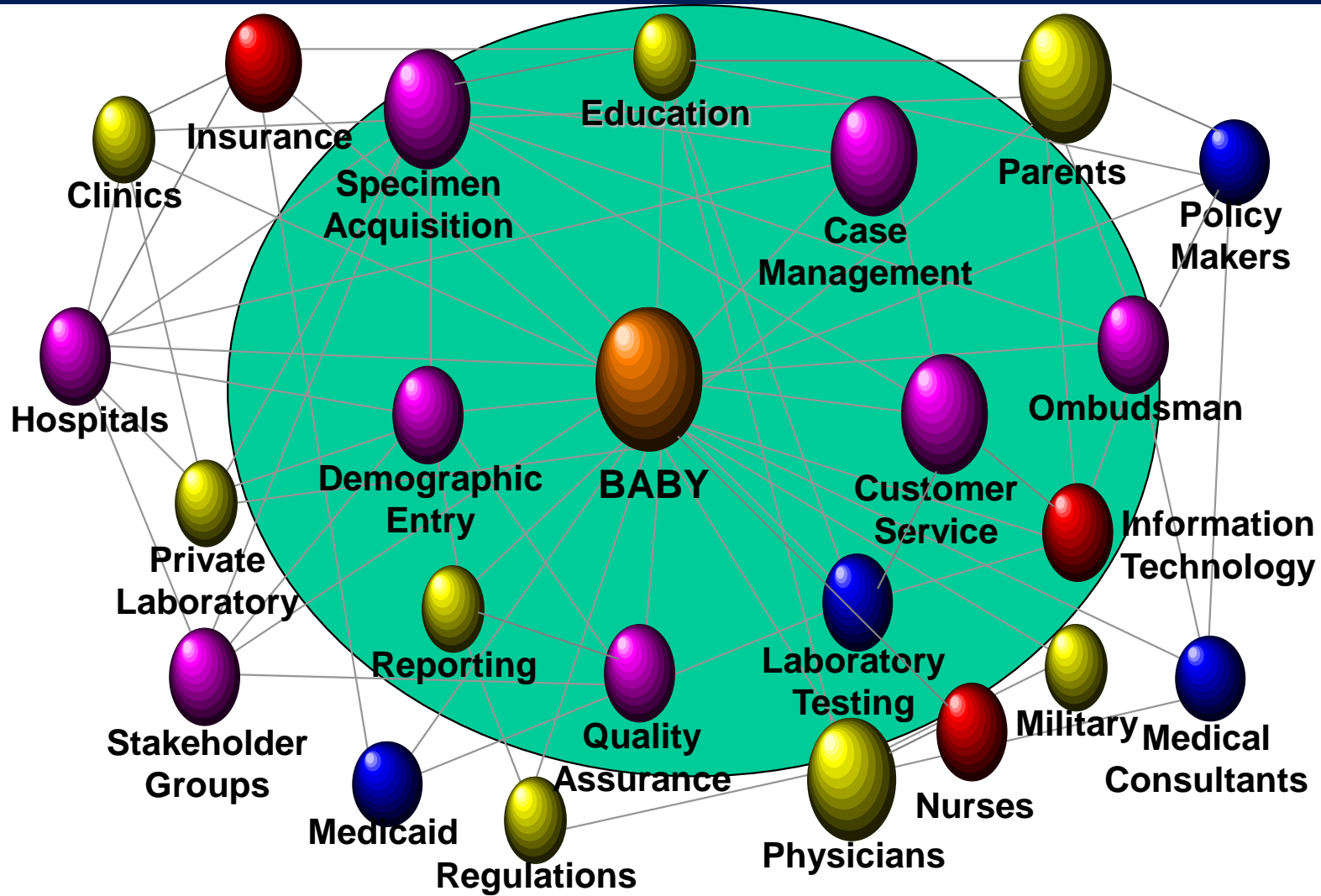
# Overview of Newborn Screening Laboratory Processes and Quality Management

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# Newborn Screening System





## Newborn Screening: Toward a Uniform Screening Panel and System

### Contents

<b>EXECUTIVE SUMMARY</b> .....	1s
<b>MAIN REPORT</b> .....	12s
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<i>INTRODUCTION</i> .....	14s

#### SECTION I

Developing a Uniform Screening Panel

## Key Components of Newborn Screening

Education (throughout the process)

Screening, including specimen collection and testing

Follow-up and result reporting

Diagnostic confirmation

Management

Program evaluation and Continuous Quality Improvement



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### SECTION I

Developing a Uniform Screening Panel

**Total quality management should be applied to newborn screening programs.**

As with any programmatic effort, improvements result from careful and continuous monitoring of key steps in the process, the assessment of that information, and the introduction of changes that continuously improve program performance. Uniform and consistent monitoring of system quality indicators can provide information about the relative performance of screening programs.

- **Quality Assurance (QA)** – all the planned and systematic activities implemented within the quality system, and demonstrated as needed, to provide adequate confidence that an entity will fulfill requirements for quality. **NOTE:** Quality assurance may be said to comprise internal quality assurance and external quality assurance and is interrelated with quality control.
- **Quality Control (QC)** – the operational techniques and activities that are used to fulfill requirements for quality.
- **Quality Indicators (QI)** - a metric that gives an indication of process or output quality and can be used to make comparisons across different Programs

# Ensuring Quality



## Newborn Screening Quality Assurance Program

QUALITY CONTROL

MIDYEAR REPORT

Volume 23, No. 1

June 2012

### INTRODUCTION

The Newborn Screening Quality Assurance Program (NSQAP), Centers for Disease Control and Prevention (CDC), distributed dried-blood-spot (DBS) quality control (QC) materials for thyroxine (T4), thyroid-stimulating hormone (TSH), 17  $\alpha$ -hydroxyprogesterone (17-OHP), total galactose (TGal), immunoreactive trypsinogen (IRT), phenylalanine (Phe), leucine (Leu), methionine (Met), tyrosine (Tyr), valine (Val), citrulline (Cit), arginine (Arg), succinylacetone (SUAC), and sixteen acylcarnitines (C0, C2, C3, C3DC, C4, C5, C5DC, C5OH, C6, C8, C10, C12, C14, C16, C16OH, C18) to laboratories operating newborn screening programs and to manufacturers of screening test products. Included with each semiannual shipment of QC specimens were instructions for downloading and submitting the paperless data report forms.

This midyear report contains a summary of the QC data submitted during the first half of 2012 by state, contract, and private laboratories in the United States; international participants; and manufacturers of screening test products.

### QUALITY CONTROL MATERIALS

The QC specimen lots were provided as 6-month supplies of DBSs on filter paper. DBS QC lots were prepared from whole blood of 50% hematocrit. The QC materials were enriched with predetermined quantities of the selected analytes and dispensed in 100  $\mu$ L aliquots on GE Healthcare Bio-Sciences Corporation (formerly Whatman

Inc.), Westborough, MA, Grade 903; and PerkinElmer Health Sciences (formerly Ahlstrom Filtration LLC), Greenville, SC, Grade 226 filter papers.

A QC shipment for T4, TSH, or 17-OHP consisted of blood-spot materials from three lots per analyte, with each lot containing a different concentration of analyte. A QC shipment for IRT, TGal, Phe, Leu, Met, Tyr, Val, Cit, Arg, SUAC and the acylcarnitines consisted of blood-spot cards from four different lots.

The QC materials were supplied for use as external controls in quantities sufficient to maintain continuity and transcend changes in production lots of routinely used method- or kit-control materials. The external QC materials were intended to supplement the participants' method- or kit-control materials at periodic intervals and to allow participants to monitor the long-term stability of their assays. The QC materials should not be used as routine daily QCs.

### PARTICIPANTS' RESULTS

For this midyear report, we compiled the data that each participant reported from five analytic runs of specimens from each QC lot and calculated mean values and standard deviations from these data. Data values outside the 95% confidence interval for each QC lot were not included in the computations. We could not include qualitative data, data submitted as inequalities or ranges, data submitted in unidentified units, or data from more than five analytic runs per specimen lot per participant. Some participants submitted results in units other than those requested on the data-report forms. To ensure that all results are appropriately entered in the CDC database, participants

---- QC DATA ----  
see pages 3-33



Clinical  
Laboratory  
Improvement  
Amendments



CCDIAPHIL

This program is organized by the Centers for Disease Control and Prevention (CDC) and the Association of Public Health Laboratories (APHL).

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Centers for Disease Control and Prevention

**MMWR**

Morbidity and Mortality Weekly Report

Recommendations and Reports / Vol. 61 / No. 2

April 6, 2012

# Good Laboratory Practices for Biochemical Genetic Testing and Newborn Screening for Inherited Metabolic Disorders

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# Preanalytic

Test selection and ordering

Specimen collection, handling, and delivery

Specimen receipt and accessioning

# Analytic

Specimen preparation

Test performance

Monitoring and verification of test accuracy and results

Documenting test findings

# Postanalytic

Reporting test results

Turn around time

Verifying electronic data transfers

Records and specimen retention



# Establishing a New Test



- Verify manufacturer's
  - Accuracy
  - Precision
  - Reportable range
  - Reference intervals



- Establish assay's
  - Accuracy
  - Precision
  - Analytical sensitivity
  - Analytical specificity
  - Reportable range
  - Reference intervals
  - Other performance characteristics

BABY'S LAST NAME (PRINT)

SN 13100001

DO NOT WRITE IN THIS AREA!

Birth Date Date of Sample Type of Feeding  
 Breast  HAL/TPN  
 Bottle  Other

Antibiotic?  Yes  No  
BABY'S MEDICAL RECORD NO.

Birth Time  am  pm Sample Time  am  pm Multiple Birth?  Yes  No  
If Yes, A, B, C, etc.:

Meconium Ileus?  Yes  No  
Remarks

Gender  M  F Birthweight gms

Transfusion PRIOR to sample collection? If Yes, give date and time:  No  Yes-

Gestational Age wks

MOTHER'S NAME (LAST, FIRST) (PRINT)

Mother's Age

Mother's Telephone No.

Address Apt. #

Mother's Race  
1  White 5  Native Hawaiian or Other Pacific Islander  
2  Black or African American 8  Other  
3  Asian 4  American Indian/Alaskan Native

Mother's Hispanic Origin  Yes  No

Collector's Initials / Date:

City, State, Zip

Mother's SSN (Last 4 digits)

HOSPITAL NAME AND ADDRESS

BABY'S PHYSICIAN NAME AND ADDRESS

Telephone No.

Telephone No.

IBM-1 AUG 13

SPECIMEN SUBMITTED BY:  Hospital  Baby's Physician

H5704

NJDOH/NBS LAB COPY

New Jersey Department of Health

INITIAL NEWBORN SCREENING REQUEST



SN 13100001



Completely fill 5 circles with blood.



- CLSI -Clinical and Laboratory Standards Institute

[www.clsi.org](http://www.clsi.org)

- LA04-A5 - Blood Collection on Filter Paper for Newborn Screening Programs; Approved Standard - Fifth Edition
- LA04-A5-DVD - Making a Difference Through Newborn Screening: Blood Collection on Filter Paper

LA4-A5  
Vol. 27 No. 20  
Replaces LA4-A4  
Vol. 23 No. 21

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## Blood Collection on Filter Paper for Newborn Screening Programs; Approved Standard—Fifth Edition

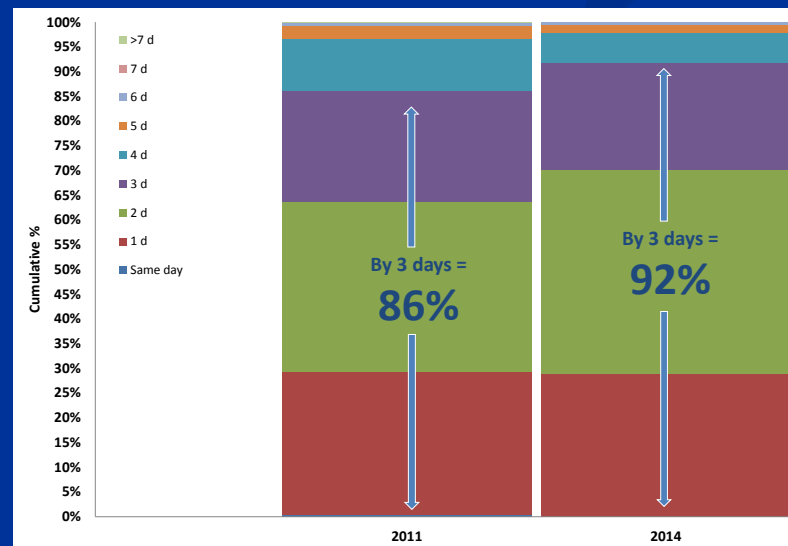
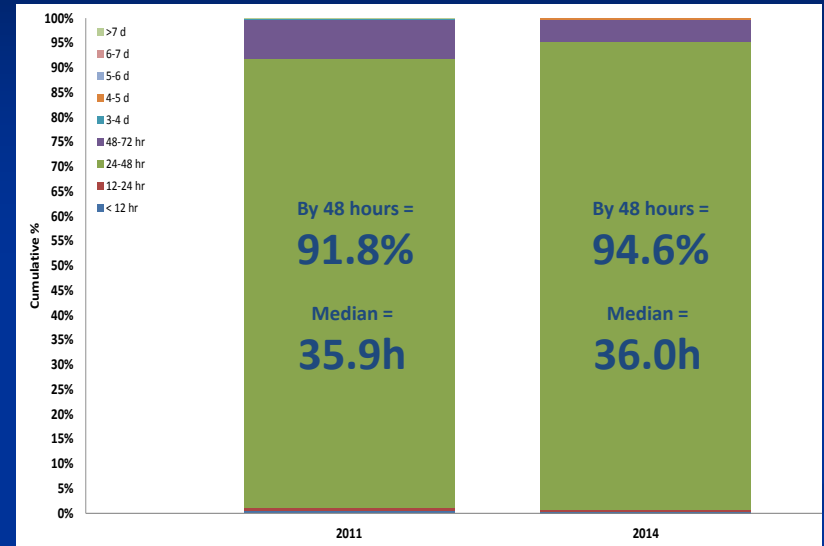
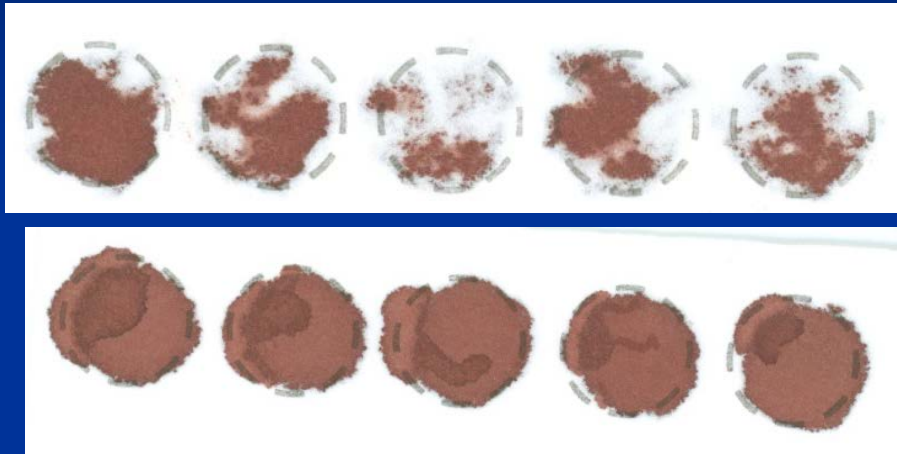
This document addresses the issues associated with specimen collection, the filter paper collection device, and the application of blood to filter paper, and provides uniform techniques for collecting the best possible specimen for use in newborn screening programs.

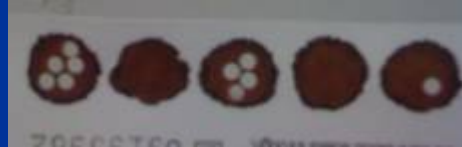
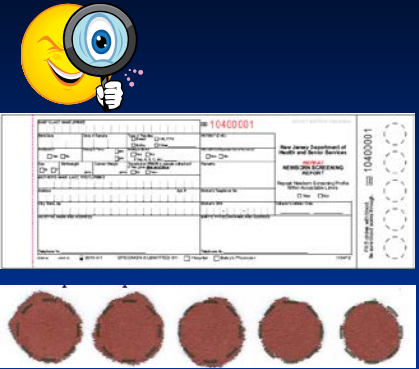
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A standard for global application developed through the Clinical and Laboratory Standards Institute consensus process.



# Preanalytic Quality







Galactosemia

- GALT
- TGAL

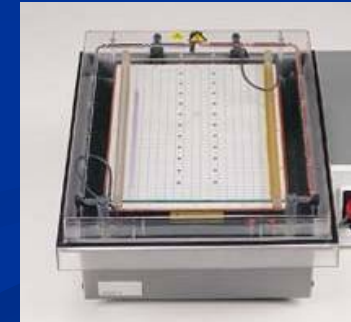
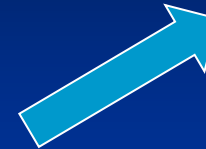
Biotinidase Deficiency

- BIO

Amino Acid Disorders

Fatty Acid Disorders

Organic Acid Disorders



Hemoglobinopathies



Cystic Fibrosis

- IRT

Congenital Adrenal Hyperplasia

- 17OHP

Congenital Hypothyroidism

- T4
- TSH



Severe Combined Immunodeficiency



# Rejecting assays is not the purpose of quality control



Process must detect immediate errors caused by test failure

# Analytic Quality

## ■ Material

- Dried blood spots
  - In-house
  - Commercial vendor
    - Kit
    - Non-kit
    - CDC NSQAP

## ■ Levels

- Decision points
  - WNL
  - Abnormal

## ■ Establishing laboratory range

- Replicates  $\geq 20$  observations
- Instrument to Instrument

## ■ Frequency

- $\geq 2$  control materials per assay

## ■ Acceptance Criteria

- Westgard rules
- Patients

## ■ Monitor

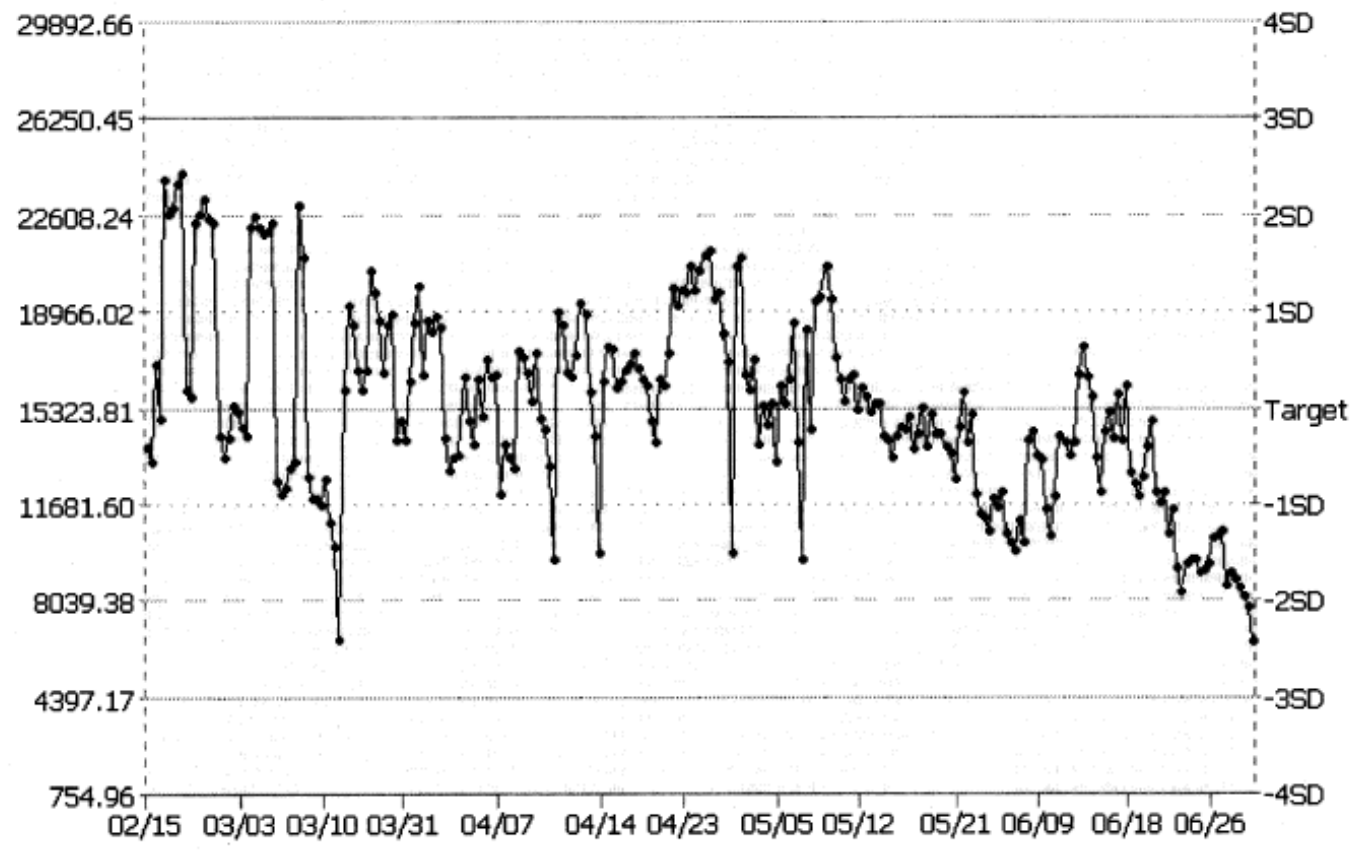
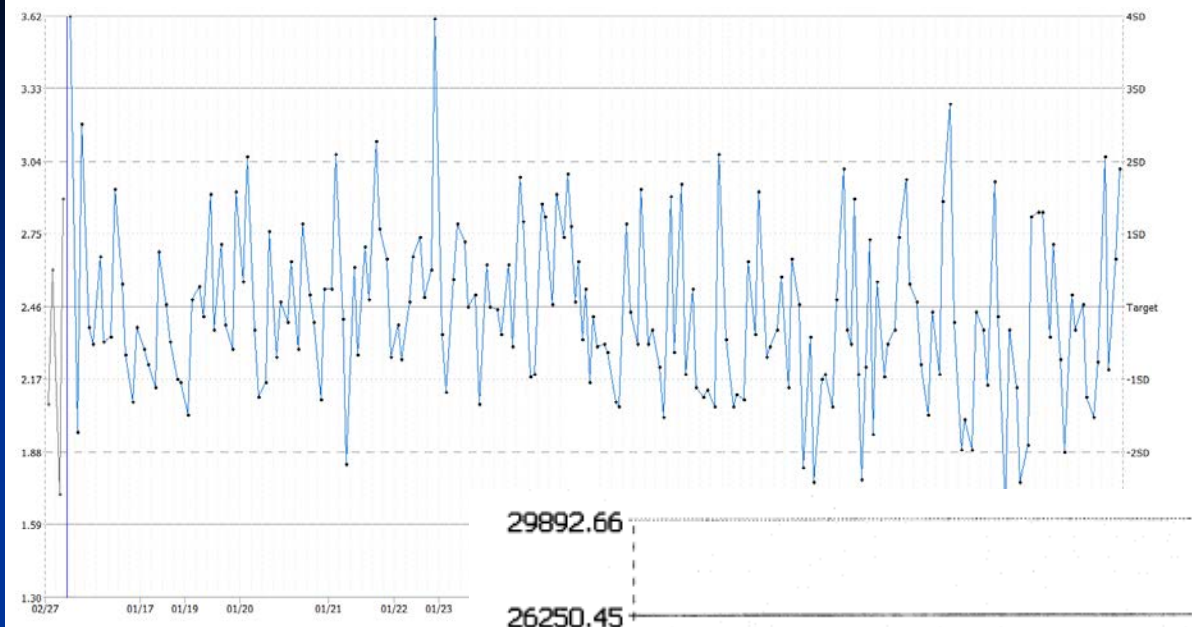
- Plate
- Instrument to Instrument
- Trends
- Shifts



# Monitor & Corrective Action

Sample	2015035P1-LOW1	2015035P1-LOW2	Range	2015035P1-HIGH1	2015035P1-HIGH2	Range
C0	139.46	157.12	(123.52 - 181.60)	358.85	342.54	(302.32 - 424.90)
C2	32.49	35.83	(29.79 - 42.99)	73.68	73.62	(61.97 - 90.35)
C3	8.33	9.44	(7.46 - 11.60)	23.56	22.56	(19.13 - 29.75)
C4	1.68	2.65	(1.39 - 4.09)	7.73	5.95	(3.57 - 10.23)
C4 MRM	2.31	2.63	(2.21 - 3.35)	6.68	6.27	(5.59 - 8.77)
C5	1.23	1.06	(0.50 - 1.58)	2.29	2.26	(1.35 - 3.93)
C5 MRM	0.94	1.05	(0.87 - 1.23)	2.49	2.55	(2.06 - 3.44)
C5DC	0.63	0.67	(0.48 - 0.72)	1.53	1.48	(1.24 - 1.90)
C6	0.42	0.63	(0.17 - 1.07)	2.19	1.46	(0.50 - 2.84)
C8	0.8	0.62	(0.24 - 1.08)	1.79	1.4	(0.71 - 2.99)
C10	0.55	0.49	(0.14 - 0.86)	1.28	0.61	(0.40 - 2.26)
C12	2.11	1.9	(1.08 - 2.82)	4.1	5.22	(2.98 - 7.42)
C14	1.99	2.19	(1.06 - 2.62)	3.65	5.43	(2.92 - 6.58)
C16	10.86	10.86	(8.09 - 17.09)	24.34	27.81	(20.27 - 40.85)
C16 MRM	11.7	13.62	(10.62 - 14.70)	31.59	29.63	(25.54 - 37.42)
C18	2.26	2.35	(1.59 - 3.33)	5.2	5.98	(2.66 - 7.40)
C18 MRM	2.36	2.56	(2.02 - 2.98)	4.87	4.67	(4.06 - 5.98)
CIT	161.82	174.12	(128.98 - 191.02)	526.25	509.94	(416.21 - 629.45)
LEU	379.78	575.87	(363.63 - 613.17)	1614.49	1485.48	(1010.70 - 1953.48)
MET	94.2	109.52	(54.32 - 112.76)	321.77	253.68	(191.79 - 392.19)
PHE	192.89	227.81	(167.78 - 254.48)	539.97	656.16	(491.07 - 803.97)
TYR	281.85	391.81	(215.61 - 389.97)	1107.64	999.54	(702.05 - 1294.73)
ARG	6.88	6.33	(3.36 - 7.08)	5.61	5.1	(3.15 - 7.71)

LC635039)



# SCID Cutoff Adjustment

>37 weeks	BORD	PRE	Total Abnormal
June 30, 2014 to Oct 7, 2014 26,835 specimens	89 (0.33%)	14 (0.05%)	103 (0.38%)
Oct 8, 2014 to Jan 31, 2015 32,082 specimens	13 (0.04%)	3 (0.009%)	16 (0.05%)

1 Classic SCID, 1 ADA SCID, 1 Leaky SCID, 5 idiopathic T-cell lymphopenia

# Proficiency Testing

- External
- Specimen exchange
- Internal



Department of Health and Human Services  
Centers for Disease Control and Prevention

[CDC en Español](#)

## Newborn Screening Quality Assurance Program

**PROFICIENCY TESTING**

Quarterly Report

This program is cosponsored by the Centers for Disease Control and Prevention and the Association of Public Health Laboratories.  
Volume 27, No. 4

November 2014

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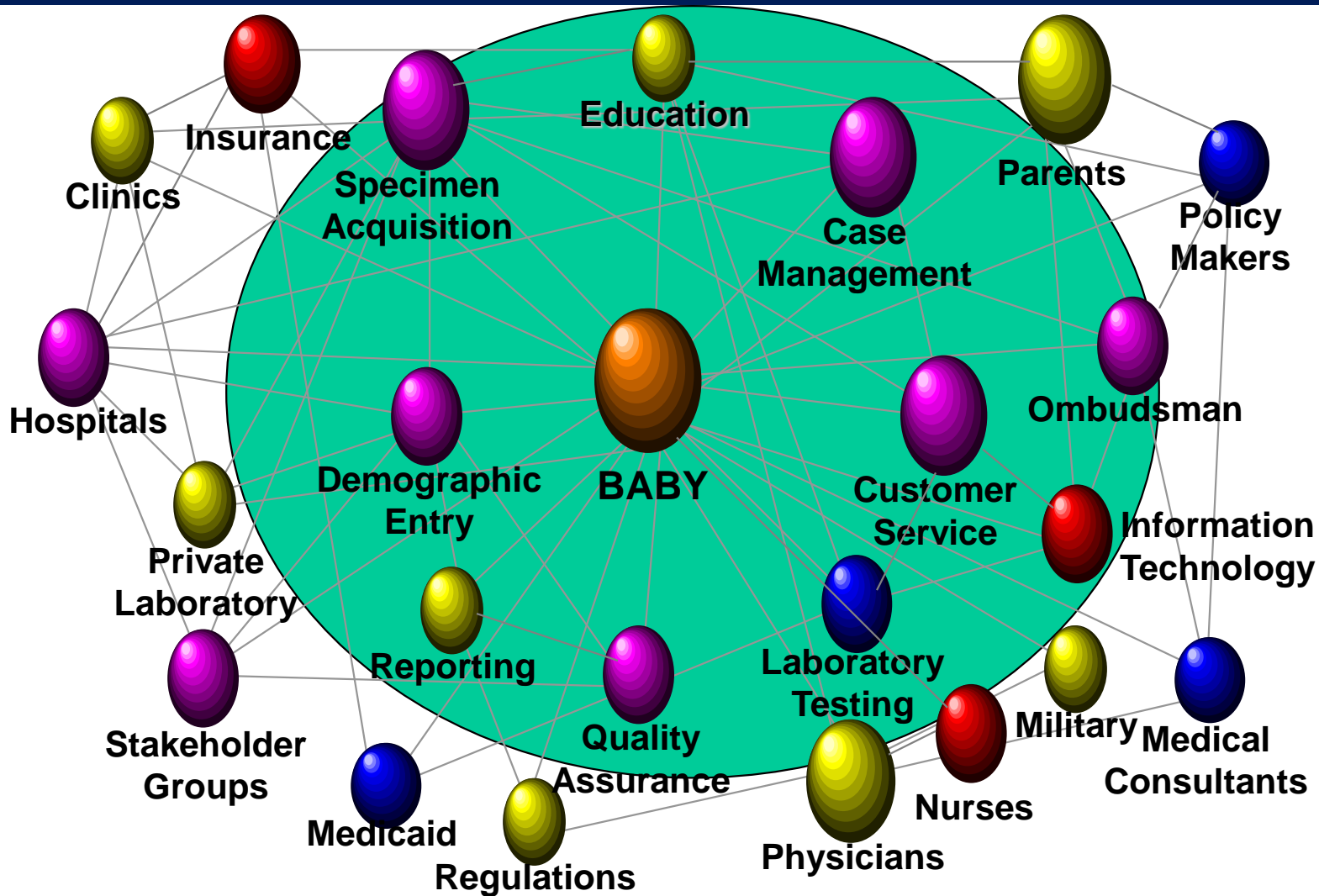
Reporting test results


Turn around time

Verifying electronic data transfers

Records and specimen retention

# Newborn Screening System





**Saving Babies  
for 50 Years  
1964 – 2014**

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State of New Jersey  
**NEWBORN  
SCREENING**